

HARNESSING GOLD NANOPARTICLES FOR TARGETED DRUG DELIVERY AND CANCER TREATMENT

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ABSTRACT

Gold nanoparticles (Au NPs) have emerged as promising radiosensitizers in various biomedical applications, including drug delivery and cancer therapy. Their unique properties, such as size, composition, morphology, and surface chemistry, can be precisely controlled through advanced synthesis and fabrication methods. Au NPs can act as contrast agents and dose enhancers in image-guided nanoparticle-enhanced radiotherapy using kilovoltage cone-beam computed tomography. Surface functionalization of Au NPs with various ligands, such as PEG, ssDNA, antibodies, peptides, drugs, fluorescence markers, and siRNA, enables them to perform multiple biomedical functions simultaneously at the molecular or cellular level. This review highlights various synthesis methods for Au NPs, including the colloidal method, galvanic replacement, and the Brust-Schiffrin method. The properties and applications of Au NPs in drug delivery and cancer therapy, such as plasmid DNA vector delivery, RNA delivery, and gold nanoparticle-based therapy, are explored. However, the potential toxicity and health impact of Au NPs need to be thoroughly investigated before clinical implementation. The review also discusses the challenges associated with drug administration and the role of biomaterials in improving medical technologies through increased control of immune signals. The synthesis and properties of Au NPs, including their core size, stabilizing agents, and surface modifications, are also discussed in detail.

KEYWORDS: Gold Nanoparticles; Drug Delivery; Cancer Therapy; Surface Plasmon Resonance; Tumor Targeting; Nanomedicine.

1. INTRODUCTION

Gold nanoparticles (Au NPs) are emerging as efficient radiosensitizers and contrast agents in biomedical and cancer therapy. Leveraging kilovoltage cone-beam computed tomography, Au NPs enhance radiotherapy by improving dosage delivery and imaging capabilities.^[1,2] Recent advances in nanomaterial synthesis have enabled precise control over particle characteristics such as size, shape, and surface chemistry, which enhances stability and functionality.^[3]

Biocompatible coatings and functional ligands can be applied to Au NPs, allowing them to serve multiple roles including drug delivery, cancer therapy, and multimodal imaging. Au NPs offer unique advantages due to their optical properties, ease of synthesis, and chemical stability, making them suitable for a range of biomedical applications. They have been utilized in gold nanoparticle-based therapies, RNA and DNA delivery, and as contrast agents in imaging.^[4-6]

Despite their potential, it is crucial to thoroughly investigate their toxicity and health effects before widespread clinical use.^[8] In addition to Au NPs, various other nanomaterials such as liposomes, carbon nanotubes, and quantum dots are employed in biomedical fields.^[9] Au NPs, often referred to as “potable gold,” are valued for their high x-ray absorption and localized surface plasmon resonance.^[16,17] Their utility in drug delivery can be influenced by factors like vascular characteristics and immune response, which affect drug accumulation and effectiveness.^[19] Advances in surface coating technology now allow for extensive functionalization of Au NPs, enhancing their role as therapeutic agents, molecular sensors, and delivery systems.^[20] Other nanomaterials like liposomes, carbon nanotubes, polymeric micelles, graphene, and quantum dots are also frequently utilized in biomedical applications.^[10] As NP-based technologies advance, human exposure to manufactured nanoparticles becomes more inevitable, making their benefits and characteristics

increasingly relevant.^[11,13] NPs, with their small size, high chemical reactivity, and large surface area, continue to be a focus of research for drug delivery and other applications.^[14] The historical use of Au NPs as “potable

gold” underscores their long-standing significance, and ongoing advancements are likely to expand their applications further.^[15,16]

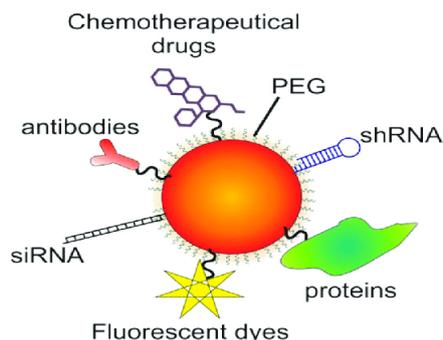


Figure 1: Representation of an Au NP for theranostics.

2. SYNTHESIS AND PROPERTIES

Gold nanoparticles (Au NPs) are commonly synthesized using the colloidal approach, which allows precise control over size, shape, and optical properties by combining a reducing agent, a metal precursor, and a stabilizing agent.^[21,22]

This method produces various nanostructures, including spheres, nanorods, and nanocages.^[23] Recent developments have introduced hollow Au NPs via galvanic replacement.^[24] and a one-pot method by Brust and Schiffrin for creating monolayer-protected Au NPs using tetrachloroaurate and sodium borohydride or citrate.^[25] The core size, ranging from 1.5 to 6 nm, is influenced by the ligand used, with larger sizes achievable through citrate reduction or ripening processes.^[26] The place exchange reaction is another technique for modifying ligand compositions on Au NPs.^[27] In medicinal applications, the bottom-up approach is favored, often using sodium citrate or sodium borohydride for reduction, and stabilizing agents like polyethyleneimine (PEI) are employed to prevent agglomeration.^[21,27] PEI facilitates nucleic acid delivery due to its amine groups.^[29] PEGylation, using

polyethylene glycol (PEG), reduces the early removal of NPs from the bloodstream.^[31] while biomimetic modifications and conditional PEG removal offer alternative surface modification techniques.^[32,33]

Different synthetic methods for Au and other metallic NPs include sol-gel micro reactors, acidic reduction, γ -irradiation, and biosynthesis.^[21] Adjusting reducing agents or chloride ions in citrate reduction can vary Au NP sizes between 19 and 47 nm.^[34] Acidic stabilization and polymeric NPs, such as glutathione-capped Au NPs, enable further size control.^[35,36] Seed-mediated synthesis uses halides for surface passivation and growth regulation.^[37] and silver ions can alter particle morphologies.^[38] Gold silica nanoshells, made by seed-mediated development, are used in imaging and targeted therapy, while gold nanorods offer adjustable near-infrared absorption with high coefficients.^[39,40] Gold nanocages and hollow Au NPs are advantageous for photothermal applications due to their unique structures.^[41] Emerging technologies in Au NP synthesis, including stimuli-responsive designs, are enhancing their applications in cancer therapy and drug delivery.^[42]

Type of NPs	Synthetic Method	NP Size (nm)
Au	Sol-gel micro reactors	5–50
Au, Ag, Pd, Pt	PVD into liquid substrate	2–10
Pd, Pt	Reduction in acidic environment	3–40
Au	Reduction process	2–40
Au, Cu	γ -Irradiation	3–30
Cu	pH control of Cu complexes	48–150
Au, Ag, Pd	Biosynthesis	9–25
Ag	Wet chemistry	20–60

3. CELLULAR UPTAKE OF GOLD NANOPARTICLE AND CYTOTOXICITY

Nanoparticles, with sizes comparable to biomolecules, can be tailored for specific biological interactions. Gold nanoparticles (Au NPs) modified with simian virus 40 (SV40) nuclear localization signals are used for targeted nuclear delivery.^[43] Techniques such as using Tat

peptides or mycobacterium-coated Au NPs enable entry into the cell nucleus and cytoplasm, respectively.^[44]

Surface functionalization is crucial for cellular uptake, influenced by ligand density and molecular weight, while Au NP size has minimal effect on uptake.^[45] Shape also affects cellular absorption, with triangular Au NPs being

more effective in RAW264.7 cells than rod-shaped or star-shaped NPs.^[46] Studies comparing hollow Au nanocages, nanorods, and plasmons with siRNA reveal that hollow Au nanoshells load more siRNA and have better gene silencing efficiency.^[47] Genotoxicity assessments of Au NPs ranging from 5 to 50 nm show that smaller particles can induce DNA damage and clastogenic

effects, especially at 5 nm.^[48] Toxicity varies with size, cell type, and tissue interaction, with smaller Au NPs (1-5 nm) exhibiting higher toxicity.^[50] Polyethylene glycol (PEG) coating enhances transfection efficacy and reduces toxicity.^[51,52] PEG-coated Au NPs show prolonged blood circulation and targeted delivery to the liver and spleen.^[54]

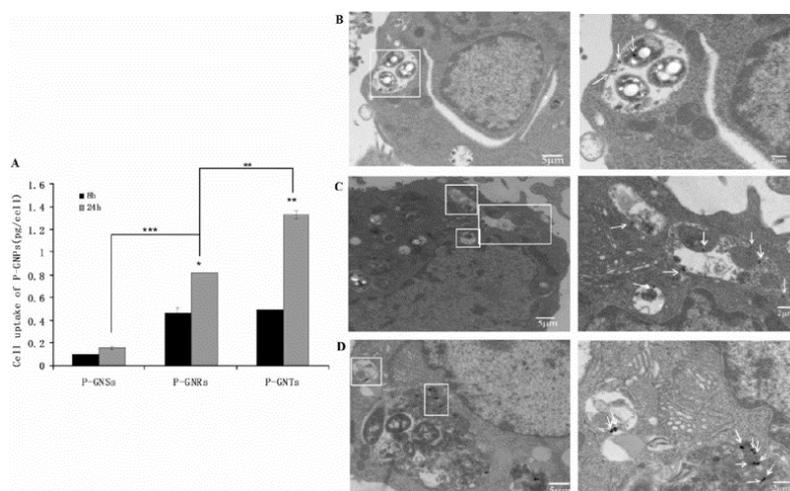


Figure 2: Cellular uptake of P-GNPs (A) and transmission electron microscopy images of RAW 264.7 after 24 h of incubation with P-GNSs (B), P-GNRs (C), P-GNTs (D). Data represent mean \pm SEM (n = 3). Statistical significance is represented by * $p < 0.05$, ** $p < 0.01$, * $p < 0.001$.^[46] Reproduced from reference.^[46] with Springer Nature under the Creative Commons Attribution 4.0 International License.**

Gold nanoparticles are also utilized to overcome multidrug resistance (MDR) in cancer therapy. Au NPs quench fluorescence efficiently, making them suitable for fluorescent nanoprobe applications. For instance, doxorubicin tethered to Au NPs via PEG improves drug delivery and efficacy against P-gp mediated drug resistance.^[55] Targeted delivery of siRNA using unimer polyion complex Au NPs and cyclic Arg-Gly-Asp ligands enhances gene silencing in cancer cells.^[56] In liver cells, green Au NPs induce oxidative stress and DNA damage, with higher sensitivity observed in cancer cells.^[57] Plasma treatment can reduce Au NP size and enhance their cytotoxicity against cancer cells.^[58]

Overall, Au NPs exhibit stable chemistry, minimal toxicity, and significant potential for biomedical applications. Further research is needed to optimize their uptake, toxicity, and therapeutic efficacy.

4. GOLD NANOPARTICLE BASED DRUG DELIVERY

Nanomedicine, particularly with gold nanoparticles (Au NPs), shows promising clinical efficacy with enhanced therapeutic results and reduced toxicity. Although Au NP-based nano-drugs are not yet officially approved, research is actively exploring their potential, especially in cancer treatment and tumor targeting.^[59] Au NPs are effective carriers for a range of drugs, including peptides, pDNAs, proteins, small siRNAs, and chemotherapeutic agents. Their surface

can be modified with functional groups like carboxyl, amine, or thiol to improve drug delivery and protect the drug from enzymatic degradation.^[60] Fucoidan, a bioactive polysaccharide, is used to synthesize Au NPs, enhancing biocompatibility and reducing toxicity compared to conventional methods.^[61] pH-sensitive medications, such as Morin encapsulated in Au NPs, show effectiveness in targeting acidic tumor environments and enhancing tumor apoptosis in mouse models.^[62] For bacterial infections, Au NPs improve drug delivery and effectiveness, especially when combined with antibiotics like gentamicin sulfate, which often suffers from poor membrane permeability and high solubility.^[64] Innovative delivery systems use Au NPs to encapsulate multiple drug molecules, allowing effective penetration into cells.^[65] Additionally, Au NPs combined with everolimus have shown promise in treating conditions like Bronchiolitis obliterans syndrome.^[66] Cyclic peptide-capped Au NPs offer enhanced drug delivery due to their ability to penetrate cell membranes and deliver therapeutic agents more efficiently.^[67] Recent advancements also include Au NP-based systems for fluorescence imaging and targeted drug delivery. For instance, Au NPs combined with platinum drugs and aminoanthraquinone demonstrate effective DNA binding and imaging capabilities.^[69] Au NPs are being explored for intraocular drug delivery, showing potential in treating retinal conditions.^[70] Moreover, cell membrane-coated Au NPs, such as those with platelet membranes, offer targeted cancer therapy

options.^[71], and doxorubicin-loaded Au NPs show promise in prostate cancer treatment.^[72] Testing in leukemia cells has also demonstrated encouraging results.^[73]

4. 1. Plasmid Deoxynucleic Acids Vector (pDNAs)

Delivery Gold nanoparticles (Au NPs) functionalized with DNA are increasingly used in drug delivery and biosensing due to their versatility and efficiency. The DNA-Au NP conjugate has shown effectiveness as a nano-carrier for drugs and genes, overcoming challenges such as nuclease degradation and blood serum interference.^[74] PEGylation of DNA strands attached to Au NPs can prevent random protein adsorption and protect against DNase I degradation, thereby enhancing cellular uptake and efficiency.^[75] Recent advances have focused on functionalizing Au NPs with oligonucleotides through thiol modification, enabling the creation of complex nanostructures such as tetramers, trimers, and dimers. These structures offer stability and can perform multiplexed functions in biological settings, including targeting mRNA and delivering anticancer drugs.^[76] Additionally, amphiphilic coatings on Au NPs, such as pyridinium amphiphiles, improve plasmid DNA delivery by influencing NP shape and size, which affects cellular uptake and DNA compaction.^[77] Nanogels containing polymeric Au NPs offer a flexible delivery system, capable of encapsulating and releasing drugs with reduced cytotoxicity. Poly(N-isopropylacrylamide)-based nanogels, for example, provide biocompatibility and temperature-responsive properties, allowing for controlled NP size changes.^{[78][79]} Liposomes, particularly those functionalized with DOTAP lipid, are also effective in gene and drug delivery, though PEG elimination from liposomes can be crucial for their efficacy.^[80] Further research has demonstrated the potential of Au NPs in gene targeting, such as blocking specific genes in cancer cells and using snake venom-derived peptides like cotamine to facilitate DNA delivery.^{[81][82]} Au NPs functionalized with Y-type DNA have been developed for detecting telomerase and releasing anticancer agents in cancer cells, providing a novel approach for cancer treatment.^[83] Additionally, NIR-sensitive nanoparticles that integrate PEG, DNA strands, and Au nanorods have been explored for thermo-chemotherapy, combining heat conversion with therapeutic DNA delivery.^[84]

4. 2. Ribonucleic Acids (RNAs) Delivery

Recent advancements have highlighted the potential of gold nanoparticles (Au NPs) in RNA delivery for HIV treatment and other applications. One approach involved PEGylated Au NPs with a covalent bond to thiol-modified oligoribonucleotides via a cleavable linker, N-succinimidyl 3-(2-puridyldithio) propionate (SPDP). The Au NPs were further coated with polyethyleneimine to enhance endosomal escape and cellular uptake. Anti-CD4 cyclic targeting peptides were attached to the Au NPs to improve selectivity and

uptake in target cells. Each lymphocyte took up about 45,000 RNA strands, showing potential for HIV therapy despite no antiviral activity being observed.^[85] RNA interference (RNAi) for gene and cancer therapy has shown promise with functionalized Au nanorods used to deliver stable hairpin RNA. This approach effectively targets human brain cancer cells, utilizing disulfide-cross-linked cleavage for endosomal escape. The high intracellular glutathione levels enable rapid RNA release, and PEGylation ensures stability and prolonged circulation, enhancing tumor accumulation and gene silencing effectiveness in brain cells. Encouraging results were observed in tumor-bearing mice.^[86] Addressing the challenge of delivering drugs across the blood-brain barrier for glioblastoma, researchers explored nose-to-brain direct transport using gold-iron oxide nanoparticles. This approach incorporates microRNA for glioblastoma therapy and temozolomide delivery. The technique showed promising results in animal models, indicating its potential for clinical application.^[87] In the context of triple-negative breast cancer, multilayered NPs containing metastasis suppressor microRNA (miR780) were designed for targeted delivery to reduce lung metastases. In vivo experiments demonstrated the efficacy of this approach, suggesting it could improve clinical outcomes for this aggressive cancer type.^[88] For ovarian cancer, Au NPs have been used to deliver DNA, specifically examining the antitumor effects of DOX-DNA-Au NPs across three ovarian cancer cell lines: SK-OV-3, HEY A8, and A2780. The results from the EZ-Cytox cell viability assay indicated that DOX-DNA-Au NPs exhibit significant activity and could be a viable treatment option.^[89]

4.3. Small Interfering Ribonucleic Acids (siRNAs) Delivery

Small interfering RNAs (siRNAs) hold significant therapeutic promise, particularly in cancer treatment. Their efficacy is hindered by challenges such as instability and low cellular uptake. Gold nanoparticles (Au NPs) have emerged as effective vectors to address these issues. In glioblastoma therapy, Au NPs encapsulated with siRNA and modified with arginine-glycine-aspartic (RGD) peptides demonstrated effective gene silencing and high transfection efficiency in U87MG cells. This was confirmed through flow cytometry, protein expression analysis, and confocal microscopy.^[91] For prostate cancer, a study utilized Au NPs coated with polyethyleneimine (PEI) and conjugated with anisamide-targeting ligands to deliver siRNA targeting the RelA gene. This approach protected siRNA from degradation and enhanced gene silencing in PC3 prostate cancer cells, showing potential for therapeutic use.^[92] In melanoma, Au NPs co-delivering anti-STAT3 siRNA and imatinib mesylate achieved a significant reduction in tumor volume and weight, highlighting a promising strategy for targeting the STAT3 pathway.^[93] For breast cancer, particularly involving cancer stem cells (CSCs), Au

NPs assembled with a polymer complex and targeted with specific ligands delivered siRNA effectively, resulting in strong cellular uptake and significant gene knockdown in CSC-rich cultures. This approach shows potential for improving treatment outcomes by targeting CSCs.^[94] Additionally, Au NPs coated with HIV-1 TAT peptides were developed to deliver siRNA targeting the ROR1 antigen in breast cancer. This method demonstrated high cellular uptake and efficient gene transfection with low cytotoxicity, indicating its potential for treating invasive breast cancer.^[95] Overall, Au NPs offer advantages in drug delivery, including longer circulation half-life, better biocompatibility, and increased cellular uptake. Future research will focus on understanding their cytotoxicity and interactions with healthy cells.

5. GOLD NANOPARTICLE BASED THEORY

Photothermal therapy (PTT) represents a promising approach in cancer treatment, leveraging the ability of nanoparticles to convert light into heat for targeted tumor destruction. Gold nanoparticles (Au NPs) are particularly effective for PTT due to their excellent biocompatibility, ease of functionalization, small size for tumor penetration, and their efficient conversion of light, especially near-infrared (NIR) light, into heat. NIR light is advantageous because it penetrates deeper into tissues compared to other wavelengths, enhancing the effectiveness of PTT.^{[96][97]} Au NPs have shown potential in various applications, including localized

heating and controlled drug delivery. Their ability to convert NIR light into heat allows for targeted hyperthermia, directly damaging or abating tumor cells.^[98] Additionally, in synergistic therapies, combining drug delivery with photothermal effects has yielded promising results in breast cancer models.^[99]

Multifunctional Au NPs, such as Au nanostars, have been designed for enhanced Raman scattering (SERS) imaging and NIR-induced photothermal therapy. These nanostars cover broad NIR absorption bands and exhibit effective SERS activity and photothermal effects, making them versatile tools for cancer diagnosis and treatment.^[100] Hybrid systems incorporating Au NPs with heat-sensitive delivery mechanisms have also been developed. For example, hollow Au NPs have been used to deliver bupivacaine, demonstrating the potential for NIR-light-activated drug release and PTT applications.^[101] Similarly, NIR-absorbing Au-Au sulfide NPs have shown enhanced tumor penetration and efficacy in photothermal cancer therapy, resulting in significant long-term tumor-free survival.^[102] In radiation therapy, Au NPs can act as dose boosters, increasing the destruction of cancer cells while minimizing damage to surrounding healthy tissue. Studies have shown that adding Au NPs to the radiation therapy can enhance dose escalation and improve treatment efficacy, with higher dose escalation factors (DER) observed in smaller tumors and with specific photon beam energies.^[1]

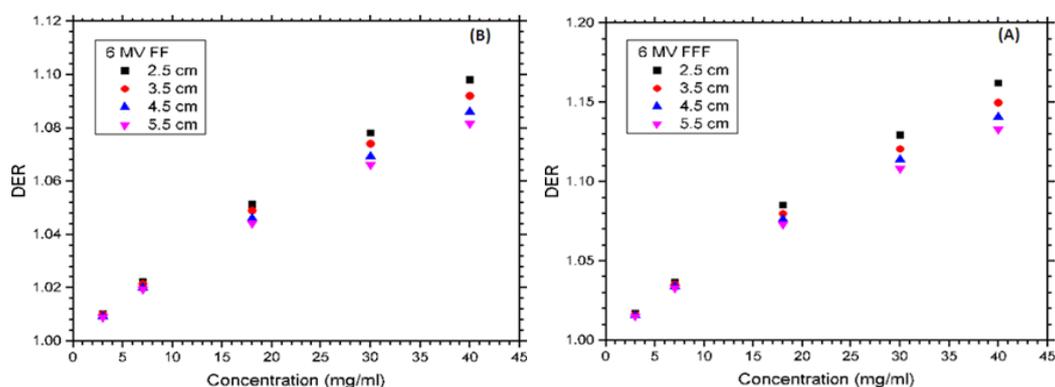


Figure 3: Relationship of the DER and Au NP concentrations, varying with different prostate sizes in the phantom using the 6 MV (A) FFF and (B) FF photon beams. Au NPs with concentrations equal to 3, 7, 18, 30, and 40 mg/mL were used. The DER was calculated as the ratio of the target dose with NP addition to the target dose without NP addition.^[1] For skin cancer radiotherapy, the addition of Au NPs improves dose deposition and image contrast, with greater DER values achieved with lower photon beam energies and thinner lesions. In summary, Au NPs offer significant advantages in cancer therapy through their role in PTT, drug delivery, and radiation dose boosting. Their ability to enhance therapeutic outcomes and reduce side effects underscores the need for further research to optimize their application in clinical settings.

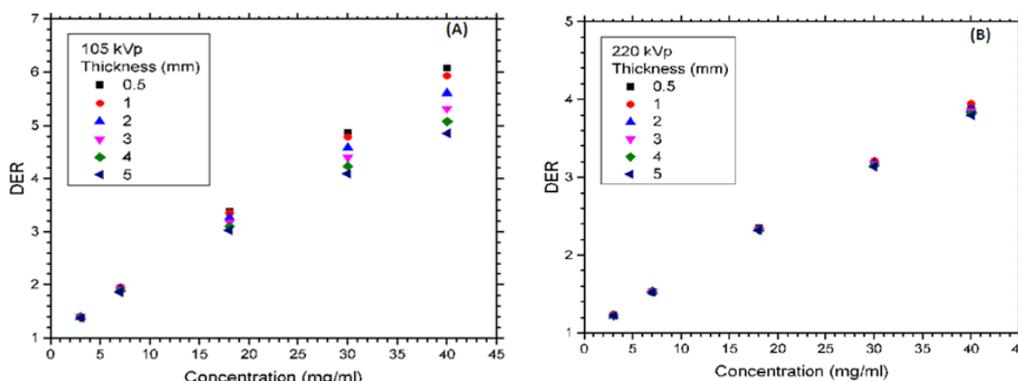


Figure 4: Relationship between the dose enhancement ratio and Au NP concentration with variation of the skin target thickness using the 105 (A) and 220 kVp (B) photon beams.^[103]

6. MOLECULAR NANOPROBES

NPs interact with light and have a tuned surface plasmon resonance, which can be detected with several imaging modalities.^[104] Au NPs have the ability to accumulate in tumor cells. Their abilities to produce photoacoustic signals and photothermal effects are also very valuable in medical and diagnostic imaging.^[105] Au NPs have optical properties that are useful in biosensors in living cells. A surface plasmon resonance scattering image for Au NPs conjugated with anti-epidermal growth factor receptor (anti-EGFR) monoclonal antibodies in a non-malignant epithelial cell line (HaCAT) and two oral epithelial cell lines malignant (HOC 313 8 and HSC clone) has been studied. Antibody-conjugated NPs bind specifically and homogeneously to the surface of cancer cells with 600% higher affinity than non-cancer cells. This produces a sharper SPR absorption band with a red-shifted maximum compared to that of the non-cancerous cell. Diffusion images generated by these antibody-conjugated gold nanoparticles are useful for diagnosis.^[106] Ultra-small gold nanoparticles less than 10 nm in diameter have shown promise in the biomedical field. Their potential applications in cancer treatment and medical imaging have not been examined. Several systems based on ultra-small gold nanoparticles are under development for use in the diagnosis and treatment of cancer. Some applications in development includes the use of ultra-small gold nanoparticles for tumor visualization and bioimaging in various fields such as magnetic resonance imaging, fluorescence imaging, photoacoustic and X-ray scattering imaging. They are also studied in tumor chemotherapy, radiotherapy and gene therapy.^[107]

7. CONCLUSION

Gold nanoparticles have high potential in cancer therapy and drug delivery applications. Although gold nanoparticles are not widely used for clinical applications, research on gold nanoparticle drug delivery, gene therapy, photothermal therapy, and radiation therapy all show promising results and are shown to be potentially viable solutions in the future. Based on the promising results obtained in the present and the progress expected in the future, it is certain that gold nanoparticles will continue to play an important role in

improving the biomedical field, especially in drug delivery and anticancer therapy. However, some limitations in the application of gold nanoparticles as nanocarriers or radiosensitizers, such as cytotoxicity, non-biodegradability and modulation of cellular responses, should not be overlooked and should be studied in detail.

REFERANCE

1. Martelli, S.; Chow, J.C.L. Dose Enhancement for the Flattening-Filter-Free and Flattening-Filter Photon Beams in Nanoparticle-Enhanced Radiotherapy: A Monte Carlo Phantom Study. *Nanomaterials*, 2020; 10: 637.[CrossRef] [PubMed].
2. Mututantri-Bastiyange, D.; Chow, J.C.L. Imaging dose of cone-beam computed tomography in nanoparticle-enhanced image-guided radiotherapy: A Monte Carlo phantom study. *AIMS Bioeng*, 2020; 7: 1–11. [CrossRef]
3. Chow, J.C.L. Monte Carlo Nanodosimetry in Gold Nanoparticle-Enhanced Radiotherapy. In *Recent Advancements and Applications in Dosimetry*; Chan, M.F., Ed.; Nova Science Publishers: New York, NY, USA, 2018; Chapter 2.
4. Jain, S.; Hirst, D.G.; O'Sullivan, J.M. Gold nanoparticles as novel agents for cancer therapy. *Br. J. Radiol.*, 2012; 85: 101–113. [CrossRef] [PubMed]
5. Chow, E.K.; Ho, D. Cancer Nanomedicine: From Drug Delivery to Imaging. *Sci. Transl. Med.*, 2013; 5: 216-4. [CrossRef] [PubMed]
6. Abdulle, A.; Chow, J.C.L. Contrast Enhancement for Portal Imaging in Nanoparticle-Enhanced Radiotherapy:A Monte Carlo Phantom Evaluation Using Flattening-Filter-Free Photon Beams. *Nanomaterials*, 2019; 9: 920.[CrossRef] [PubMed]
7. Alkilany, A.; Murphy, C. Toxicity and cellular uptake of gold nanoparticles: what we have learned so far? *J. Nanopart. Res.*, 2010; 12: 2313–2333. [CrossRef] [PubMed]
8. Jurj, A.; Braicu, C.; Pop, L.-A.; Tomuleasa, C.; Gherman, C.D.; Berindan-Neagoe, I. The new era of nanotechnology, an alternative to change cancer treatment. *Drug Des. Dev. Ther.*, 2017; 11: 2871–2890.[CrossRef]

9. Lavacchi, D.; Roviello, G.; D'Angelo, A. Tumor-Agnostic Treatment for Cancer: When How is Better than Where. *Clin. Drug Investig.*, 2020; 40: 519–527. [CrossRef]
10. Kong, F.-Y.; Zhang, J.-W.; Li, R.-F.; Wang, Z.-X.; Wang, W.-J.; Wang, W. Unique Roles of Gold Nanoparticles in Drug Delivery, Targeting and Imaging Applications. *Molecules*, 2017; 22, 1445. [CrossRef]
11. Chow, J.C.L. Application of Nanoparticle Materials in Radiation Therapy. In *Handbook of Ecomaterials*; Martinez, L.M.T., Kharissova, O.V., Kharisov, B.I., Eds.; Springer Nature: Basel, Switzerland, 2017; 150; 3661–3681.
12. Chow, J.C.L. Photon and electron interactions with gold nanoparticles: a Monte Carlo study on gold nanoparticle-enhanced radiotherapy. In *Nanobiomaterials in Medical Imaging: Applications of nanobiomaterials*; Grumezescu, A.M., Ed.; Elsevier: Amsterdam, The Netherlands, 2016; 2; 45–70.
13. Chow, J.C.L. Recent Progress of Gold Nanomaterials in Cancer Therapy. In *Handbook of Nanomaterials and Nanocomposites for Energy and Environmental Applications*; Kharissova, O.V., Torres-Martínez, L.M., Kharisov, B.I., Eds.; Springer Nature: Cham, Switzerland, 2020; 1–30.
14. Wuttke, S.; Lismont, M.; Escudero, A.; Rungtaweivoranit, B.; Parak, W.J. Positioning metal-organic framework nanoparticles within the context of drug delivery—A comparison with mesoporous silica nanoparticles and dendrimers. *Biomaterials*, 2017; 123: 172–183. [CrossRef]
15. Abbas, M.; Alqahtani, M.; AlGahtani, A.; Kessentini, A.; Loukil, H.; Parayangat, M.; Ijyas, T.; Mohammed, A.W. Validation of Nanoparticle Response to the Sound Pressure Effect during the Drug-Delivery Process. *Polymers*, 2020; 12: 186. [CrossRef] [PubMed]
16. Liz-Marzán, L.M. Gold nanoparticle research before and after the Brust–Schiffrin method. *Chem. Commun.*, 2013; 49: 16–18. [CrossRef] [PubMed]
17. Bai, X.; Wang, Y.; Song, Z.; Feng, Y.; Chen, Y.; Zhang, D.; Feng, L. The Basic Properties of Gold Nanoparticles and their Applications in Tumor Diagnosis and Treatment. *Int. J. Mol. Sci.*, 2020; 21: 2480. [CrossRef] [PubMed]
18. Dumur, F.; Dumas, E.; Mayer, C. Functionalization of Gold Nanoparticles by Inorganic Entities. *Nanomaterials*, 2020; 10: 548. [CrossRef] [PubMed]
19. Katti, K.V. Renaissance of nuclear medicine through green nanotechnology: functionalized radioactive gold nanoparticles in cancer therapy—My journey from chemistry to saving human lives. *J. Radioanal. Nucl. Chem.* 2016; 309: 5–14. [CrossRef]
20. Baptista, P.V.; Fernandes, A.R.; Figueiredo, S.; Vinhas, R.; Cordeiro, M.; Carlos, F.F.; Mendo, S. Gold nanoparticle-based theranostics: disease diagnostics and treatment using a single nanomaterial. *Nanobiosens. Dis. Diagn.*, 2015; 4: 11. [CrossRef]
21. Slepíčka, P.; Kasálková, N.S.; Siegel, J.; Kolská, Z.; Švorčík, V. Methods of Gold and Silver Nanoparticles Preparation. *Materials*, 2019; 13: 1. [CrossRef]
22. Sau, T.K.; Murphy, C. Role of ions in the colloidal synthesis of gold nanowires. *Philos. Mag.*, 2007; 87: 2143–2158. [CrossRef]
23. Austin, L.; Mackey, M.A.; Dreaden, E.C.; El-Sayed, M.A. The optical, photothermal, and facile surface chemical properties of gold and silver nanoparticles in biodiagnostics, therapy, and drug delivery. *Arch. Toxicol.*, 2014; 88: 1391–1417. [CrossRef]
24. Adams, S.; Zhang, J.Z. Unique optical properties and applications of hollow gold nanospheres (HGNs). *Co-ord. Chem. Rev.*, 2016; 320: 18–37. [CrossRef]
25. Brust, M.; Walker, M.; Bethell, D.; Schiffrin, D.J.; Whyman, R. Synthesis of thiol-derivatised gold nanoparticles in a two-phase Liquid/Liquid system. *J. Chem. Soc. Chem. Commun.*, 1994; 7: 801. [CrossRef]
26. Zhao, J.; Friedrich, B. *Synthesis of Gold Nanoparticles Via the Chemical Reduction Methods*; Shaker: Oak Park, IL, USA, 2017.
27. Kim, C.; Ghosh, P.; Rotello, V. Multimodal drug delivery using gold nanoparticles. *Nanoscale*, 2009; 1: 61–67. [CrossRef] [PubMed]
28. Kimling, J.; Maier, M.; Okenve, B.; Kotaidis, V.; Ballot, H.; Plech, A. Turkevich Method for Gold Nanoparticle Synthesis Revisited. *J. Phys. Chem. B.*, 2006; 110: 15700–15707. [CrossRef] [PubMed]
29. Wen, S.; Zheng, F.; Shen, M.; Shi, X. Synthesis of polyethyleneimine-stabilized gold nanoparticles for colorimetric sensing of heparin. *Colloids Surf. A Physicochem. Eng. Asp.*, 2013; 419: 80–86. [CrossRef]
30. Ortega-Muñoz, M.; Girón, M.D.; Salto, R.; Reyes, A.B.J.; De Jesus, S.; Lopez-Jaramillo, F.J.; Hernandez-Mateo, F.; Santoyo-Gonzalez, F. Polyethyleneimine-Coated Gold Nanoparticles: Straightforward Preparation of Efficient DNA Delivery Nanocarriers. *Chem. Asian J.*, 2016; 11: 3365–3375. [CrossRef].
31. Jokerst, J.V.; Lobovkina, T.; Zare, R.N.; Gambhir, S.S. Nanoparticle PEGylation for imaging and therapy. *Nanomedicine*, 2011; 6: 715–728. [CrossRef]
32. Amoozgar, Z.; Yeo, Y. Recent advances in stealth coating of nanoparticle drug delivery systems. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.*, 2012; 4: 219–233. [CrossRef]
33. Sengani, M.; Grumezescu, A.M.; Rajeswari, V.D. Recent trends and methodologies in gold nanoparticle synthesis—A prospective review on drug delivery aspect. *OpenNano*, 2017; 2: 37–46. [CrossRef]
34. Zhao, L.; Jiang, D.; Cai, Y.; Ji, X.; Xie, R.; Yang, W. Tuning the size of gold nanoparticles in the

- citrate reduction by chloride ions. *Nano scale*, 2012; 4: 5071. [CrossRef]
35. Cortez-Lemus, N.A.; Licea-Claverie, A.; Paraguay-Delgado, F.; Alonso-Núñez, G. Gold nanoparticles size design and control by poly(N,N 0-diethylaminoethyl methacrylate). *J. Nanomater.* 2015. [CrossRef]
36. Brinas, R.P.; Hu, M.; Qian, L.; Lymar, E.S.; Hainfeld, J.F. Gold Nanoparticle Size Controlled by Polymeric Au(I) Thiolate Precursor Size. *J. Am. Chem. Soc.*, 2008; 130: 975–982. [CrossRef] [PubMed]
37. Kasture, M.; Sastry, M.; Prasad, B.L.V. Halide ion controlled shape dependent gold nanoparticle synthesis with tryptophan as reducing agent: Enhanced fluorescent properties and white light emission. *Chem. Phys. Lett.*, 2010; 484: 271–275. [CrossRef]
38. Langille, M.R.; Personick, M.L.; Zhang, J.; Mirkin, C.A. Defining Rules for the Shape Evolution of Gold Nanoparticles. *J. Am. Chem. Soc.*, 2012; 134: 14542–14554. [CrossRef]
39. Coughlin, A.; West, J. Gold nanoshells for imaging and photothermal ablation of cancer. In *Nanomedicine*; Elsevier BV: Amsterdam, The Netherlands, 2012; 326–355.
40. Kennedy, L.; Bickford, L.R.; Lewinski, N.A.; Coughlin, A.J.; Hu, Y.; Day, E.S.; West, J.L.; Drezek, R.A. A New Era for Cancer Treatment: Gold-Nanoparticle-Mediated Thermal Therapies. *Small*, 2010; 7: 169–183. [CrossRef]
41. Sangnier, A.P.; Van De Walle, A.; Aufaure, R.; Fradet, M.; Motte, L.; Guénin, E.; Lalatonne, Y.; Wilhelm, C. Endosomal Confinement of Gold Nanospheres, Nanorods, and Nanoraspberries Governs Their Photothermal Identity and Is Beneficial for Cancer Cell Therapy. *Adv. Biosyst.*, 2020; 4: 1900284. [CrossRef] [PubMed]
42. Gellner, M.; Steinigeweg, D.; Ichilmann, S.; Salehi, M.; Schütz, M.; Kömpe, K.; Haase, M.; Schlücker, S. 3D Self-Assembled Plasmonic Superstructures of Gold Nanospheres: Synthesis and Characterization at the Single-Particle Level. *Small*, 2011; 7: 3445–3451. [CrossRef] [PubMed]
43. Wang, T.; Zhang, Z.; Gao, D.; Li, F.; Wei, H.; Liang, X.; Cui, Z.; Zhang, X.-E. Encapsulation of gold nanoparticles by simian virus 40 capsids. *Nano scale*, 2011; 3: 4275–4282. [CrossRef]
44. Mandal, D.; Maran, A.; Yaszemski, M.J.; Bolander, M.E.; Sarkar, G. Cellular uptake of gold nanoparticles directly cross-linked with carrier peptides by osteosarcoma cells. *J. Mater. Sci. Mater. Electron.*, 2008; 20: 347–350. [CrossRef]
45. Lu, H.; Su, J.; Mamdooh, R.; Li, Y.; Stenzel, M.H. Cellular Uptake of Gold Nanoparticles and Their Movement in 3D Multicellular Tumor Spheroids: Effect of Molecular Weight and Grafting Density of Poly(2-hydroxyethyl acrylate). *Macromol. Biosci.*, 2019; 20: 1900221. [CrossRef]
46. Xie, X.; Liao, J.; Shao, X.; Li, Q.; Lin, Y. The Effect of shape on Cellular Uptake of Gold Nanoparticles in the forms of Stars, Rods, and Triangles. *Sci. Rep.*, 2017; 7: 3827. [CrossRef]
47. Morgan, E.; Wupperfeld, D.; Morales, D.P.; Reich, N.O. Shape Matters: Gold Nanoparticle Shape Impacts the Biological Activity of siRNA Delivery. *Bioconj. Chem.*, 2019; 30: 853–860. [CrossRef] [PubMed]
48. Xia, Q.; Zhang, S.; Feng, Q.; Xiao, K.; Li, H.; Liu, Y. The effect of particle size on the genotoxicity of gold nanoparticles. *J. Biomed. Mater. Res. Part A*, 2016; 105: 710–719. [CrossRef] [PubMed]
49. Zhang, X.-D.; Wu, D.; Shen, X.; Liu, P.-X.; Yang, N.; Zhao, B.; Zhang, H.; Sun, Y.-M.; Zhang, L.-A.; Fan, F.-Y. Size-dependent in vivo toxicity of PEG-coated gold nanoparticles. *Int. J. Nanomed.*, 2011; 6: 2071–2081. [CrossRef] [PubMed]
50. Du, B.; Jiang, X.; Das, A.; Zhou, Q.; Yu, M.; Jin, R.; Zheng, J. Glomerular barrier behaves as an atomically precise bandpass filter in a sub-nanometer regime. *Nat. Nanotechnol.*, 2017; 12: 1096. [CrossRef].
51. Zamora-Justo, J.A.; Abrica-González, P.; Vázquez-Martínez, G.R.; Muñoz-Diosdado, A.; Balderas-López, J.A.; Ibáñez-Hernández, M. Polyethylene Glycol-Coated Gold Nanoparticles as DNA and Atorvastatin Delivery Systems and Cytotoxicity Evaluation. *J. Nanomater.*, 2019; 2019: 1–11. [CrossRef]
52. Soenen, S.J.; Manshian, B.B.; Abdelmonem, A.; Montenegro, J.-M.; Tan, S.; Balcaen, L.; Vanhaecke, F.; Brisson, A.R.; Parak, W.J.; De Smedt, S.C.; et al. The Cellular Interactions of PEGylated Gold Nanoparticles: Effect of PEGylation on Cellular Uptake and Cytotoxicity. *Part. Part. Syst. Charact.*, 2014; 31: 794–800. [CrossRef]
53. Cho, W.-S.; Cho, M.; Jeong, J.; Choi, M.; Cho, H.-Y.; Han, B.-S.; Kim, S.H.; Kim, H.O.; Lim, Y.T.; Chung, B.H.; et al. Acute toxicity and pharmacokinetics of 13 nm-sized PEG-coated gold nanoparticles. *Toxicol. Appl. Pharmacol.*, 2009; 236: 16–24. [CrossRef]
54. Lipka, J.; Semmler-Behnke, M.; Sperling, R.; Wenk, A.; Takenaka, S.; Schleh, C.; Kissel, T.; Parak, W.J.; Kreyling, W.G. Biodistribution of PEG-modified gold nanoparticles following intratracheal instillation and intravenous injection. *Biomaterials*, 2010; 31: 6574–6581. [CrossRef]
55. Wang, F.; Wang, Y.; Dou, S.; Xiong, M.-H.; Sun, T.-M.; Wang, J. Doxorubicin-Tethered Responsive Gold Nanoparticles Facilitate Intracellular Drug Delivery for Overcoming Multidrug Resistance in Cancer Cells. *ACS Nano*, 2011; 5: 3679–3692. [CrossRef]
56. Yi, Y.; Kim, H.J.; Mi, P.; Zheng, M.; Takemoto, H.; Toh, K.; Kim, B.G.; Hayashi, K.; Naito, M.; Matsumoto, Y.; et al. Targeted systemic delivery of siRNA to cervical cancer model using cyclic RGD-installed unimer polyion complex-assembled gold

- nanoparticles. *J. Control. Release*, 2016; 244: 247–256. [CrossRef]
57. Bin-Jumah, M.N.; Al-Abdan, M.; Al-Basher, G.; Alarifi, S. Molecular Mechanism of Cytotoxicity, Genotoxicity, and Anticancer Potential of Green Gold Nanoparticles on Human Liver Normal and Cancerous Cells. *Dose-Response*, 2020; 18. [CrossRef] [PubMed]
 58. Chokradjaroen, C.; Rujiravanit, R.; Theeramunkong, S.; Saito, N. Effect of electrical discharge plasma on cytotoxicity against cancer cells of N,O-carboxymethyl chitosan-stabilized gold nanoparticles. *Carbohydr. Polym.*, 2020; 237: 116-162. [CrossRef] [PubMed]
 59. Wang, W.; Wang, J.; Ding, Y. Gold nanoparticle-conjugated nanomedicine: design, construction, and structure-efficacy relationship studies. *J. Mater. Chem. B* 2020. [CrossRef] [PubMed]
 60. Jeong, E.H.; Jung, G.; Hong, C.A.; Lee, H.-J. Gold nanoparticle (AuNP)-based drug delivery and molecular imaging for biomedical applications. *Arch. Pharmacol Res.*, 2013; 37: 53–59. [CrossRef]
 61. Manivasagan, P.; Bharathiraja, S.; Bui, N.Q.; Jang, B.; Oh, Y.-O.; Lim, I.G.; Oh, J. Doxorubicin-loaded fucoidan capped gold nanoparticles for drug delivery and photoacoustic imaging. *Int. J. Boil. Macromol.*, 2016; 91: 578–588. [CrossRef]
 62. Ding, X.; Yin, C.; Zhang, W.; Sun, Y.; Zhang, Z.; Yang, E.; Sun, D.; Wang, W. Designing Aptamer-Gold Nanoparticle-Loaded pH-Sensitive Liposomes Encapsulate Morin for Treating Cancer. *Nanoscale Res. Lett.*, 2020; 15: 1–17. [CrossRef]
 63. Zharov, V.P.; Mercer, K.E.; Galitovskaya, E.N.; Smeltzer, M.S. Photothermal Nanotherapeutics and Nanodiagnostics for Selective Killing of Bacteria Targeted with Gold Nanoparticles. *Biophys. J.*, 2006; 90: 619–627. [CrossRef]
 64. Yeluri, T.; Bhosale, R.S.; Ghule, N.V.; Raynor, A.M.; Bhosale, S.V.; Bhosale, S.V. Neomycin and gentamicin detection via molecular recognition with cyclam-decorated gold nanoparticles. *Supramol. Chem.*, 2015; 27: 772–779. [CrossRef]
 65. Ahangari, A.; Salouti, M.; Heidari, Z.; Kazemizadeh, A.R.; Safari, A.A. Development of gentamicin-gold nanospheres for antimicrobial drug delivery to Staphylococcal infected foci. *Drug Deliv.*, 2013; 20: 34–39. [CrossRef]
 66. Meloni, F.; Cova, E.; Inghilleri, S.; Oggionni, T.; Magni, S.; Morosini, M.; Briganti, D.; Colombo, M.; Agozzino, M.; De Angelis, M.C.; et al. Novel Antibody-Engineered Gold Nanoparticles as Targeted Drug Delivery for Primary Mesenchymal Cells Do Not Elicit an Inflammatory Response. *J. Hear. Lung Transpl.*, 2014; 33: 166. [CrossRef]
 67. Shirazi, A.N.; Mandal, D.; Tiwari, R.K.; Guo, L.; Lu, W.; Parang, K. Cyclic Peptide-Capped Gold Nanoparticles as Drug Delivery Systems. *Mol. Pharm.*, 2012; 10: 500–511. [CrossRef] [PubMed]
 68. Borlan, R.; Tatar, A.-S.; Soritau, O.; Maniu, D.; Marc, G.; Florea, A.; Focsan, M.; Astilean, S. Design of fluorophore-loaded human serum albumin nanoparticles for specific targeting of NIH: OVCAR3 ovarian cancer cells. *Nanotechnology*, 2020; 31: 315-102. [Cross Ref].
 69. Caballero, A.B.; Cardo, L.; Claire, S.; Craig, J.S.; Hodges, N.J.; Vladyka, A.; Albrecht, T.; Rochford, L.A.; Pikramenou, Z.; Hannon, M.J. Assisted delivery of anti-tumour platinum drugs using DNA-coiling gold nanoparticles bearing lumophores and intercalators: towards a new generation of multimodal nanocarriers with enhanced action. *Chem. Sci.*, 2019; 10: 9244–9256. [CrossRef] [PubMed]
 70. De Matteis, V.; Rizzello, L. Noble Metals and Soft Bio-Inspired Nanoparticles in Retinal Diseases Treatment: A Perspective. *Cells*, 2020; 9: 679. [CrossRef]
 71. Kim, M.W.; Lee, G.; Niidome, T.; Komohara, Y.; Lee, R.; Park, Y.I. Platelet-Like Gold Nanostars for Cancer Therapy: The Ability to Treat Cancer and Evade Immune Reactions. *Front. Bioeng. Biotechnol.*, 2020; 8: 133. [CrossRef] [PubMed]
 72. Lee, C.S.; Kim, H.; Yu, J.; Yu, S.H.; Ban, S.; Oh, S.; Jeong, D.; Im, J.; Baek, M.J.; Kim, T.H. European Journal of Medicinal Chemistry Doxorubicin-loaded oligonucleotide conjugated gold nanoparticles: A promising in vivo drug delivery system for colorectal cancer therapy. *Eur. J. Med. Chem.*, 2017; 142: 416–423. [CrossRef]
 73. Li, J.; Wang, X.; Wang, C.; Chen, B.; Dai, Y.; Zhang, R.; Song, M.; Lv, G.; Fu, D. The Enhancement Effect of Gold Nanoparticles in Drug Delivery and as Biomarkers of Drug-Resistant Cancer Cells. *Chem Med Chem.*, 2007; 2: 374–378. [CrossRef]
 74. Wu, R.; Peng, H.; Zhu, J.-J.; Jiang, L.-P.; Liu, J. Attaching DNA to Gold Nanoparticles With a Protein Corona. *Front. Chem.*, 2020; 8: 121. [CrossRef]
 75. Song, L.; Guo, Y.; Roebuck, D.; Chen, C.; Yang, M.; Yang, Z.; Sreedharan, S.; Glover, C.; Thomas, J.A.; Liu, N.; et al. Terminal PEGylated DNA–Gold Nanoparticle Conjugates Offering High Resistance to Nuclease Degradation and Efficient Intracellular Delivery of DNA Binding Agents. *ACS Appl. Mater. Interfaces*, 2015; 7: 18707–18716. [CrossRef]
 76. Kyriazi, M.-E.; Giust, D.; El-Sagheer, A.H.; Lackie, P.M.; Muskens, O.L.; Brown, T.; Kanaras, A.G. Multiplexed mRNA Sensing and Combinatorial-Targeted Drug Delivery Using DNA-Gold Nanoparticle Dimers. *ACS Nano* 2018, 12: 3333–3340. [CrossRef]
 77. Kizewski, A.; Ilies, M.A. Efficient and synergetic DNA delivery with pyridinium amphiphiles–gold nanoparticle composite systems having different packing parameters. *Chem. Commun.*, 2016; 52: 60–63. [CrossRef] [PubMed]
 78. Mackova, H.; Jendelova, P. Effects of the reaction parameters on the properties of thermosensitive

- poly(N-isopropylacrylamide) microspheres prepared by precipitation and dispersion polymerization. *J. Polym. Sci. Part A Polym. Chem.*, 2005; 44: 968–982. [CrossRef]
79. Udomluck, N.; Baipaywad, P.; Lee, S.H.; Park, H. Poly(N -isopropylacrylamide)-based nanogels encapsulating gold nanoparticles for DNA delivery. *J. Control. Release*, 2015; 213: 85. [CrossRef] [PubMed]
 80. Bhuvana, M.; Dharuman, V. Influence of alkane chain lengths and head groups on tethering of liposome–gold nanoparticle on gold surface for electrochemical DNA sensing and gene delivery. *Sens. Actuators B Chem.*, 2016; 223: 157–165. [CrossRef]
 81. Karimi, S.; Fouani, M.H.; Moshaii, A.; Nikkhah, M.; Hosseinkhani, S.; Sheikhejad, R. Development of Dual Functional Nucleic Acid Delivery Nanosystem for DNA Induced Silencing of Bcl-2 Oncogene. *Int. J. Nanomed.*, 2020; 15: 1693–1708. [CrossRef] [PubMed]
 82. Hayashi, M.A.F.; Campeiro, J.D.; Porta, L.C.; Szychowski, B.; Alves, W.A.; Oliveira, E.B.; Kerkis, I.; Daniel, M.-C.; Karpel, R.L. Crotonamine Cell-Penetrating Nanocarriers: Cancer-Targeting and Potential Biotechnological and/or Medical Applications. *Methods Mol. Biol.*, 2020; 2118: 61–89. [CrossRef]
 83. Zhang, S.; Liang, K.; Li, P.; Liu, J.; Tang, B. Application of a Y-type-DNA-functionalized nanogold probe featuring specific telomerase recognition and doxorubicin release in cancer cells. *Analyst*, 2020; 145: 2152–2158. [CrossRef]
 84. Xiao, Z.; Ji, C.; Shi, J.; Pridgen, E.M.; Frieder, J.; Wu, J.; Farokhzad, O.C. DNA self-assembly of targeted near-infrared-responsive gold nanoparticles for cancer thermo-chemotherapy. *Angew. Chem. Int. Ed.*, 2012; 51: 11853–11857. [CrossRef]
 85. Parboosing, R.; Govender, T.; Maguire, G.E.M.; Kruger, H.G. Synthesis, Characterization and Biocompatibility of a Multifunctional Gold Nanoparticle System for the Delivery of Single-Stranded RNA to Lymphocytes. *S. Afr. J. Chem.*, 2018; 71: 1–14. [CrossRef]
 86. Wang, F.; Zhang, W.; Shen, Y.; Huang, Q.; Zhou, D.; Guo, S. Efficient RNA delivery by integrin-targeted glutathione responsive polyethyleneimine capped gold nanorods. *Acta Biomater.*, 2015; 23: 136–146. [CrossRef]
 87. Sukumar, U.K.; Bose, R.J.; Malhotra, M.; Babikir, H.A.; Afjei, R.; Robinson, E.; Zeng, Y.; Chang, E.; Habte, F.; Sinclair, R.; et al. Intranasal delivery of targeted polyfunctional gold-iron oxide nanoparticles loaded with therapeutic microRNAs for combined theranostic multimodality imaging and presensitization of glioblastoma to temozolomide. *Biomaterials*, 2019; 218: 119–342. [CrossRef] [PubMed]
 88. Ramchandani, D.; Lee, S.K.; Yomtoubian, S.; Han, M.S.; Tung, C.-H.; Mittal, V. Nanoparticle Delivery of miR-708 Mimetic Impairs Breast Cancer Metastasis. *Mol. Cancer Ther.*, 2019; 18: 579–591. [CrossRef] [PubMed]
 89. Lee, S.-W.; Park, S.-M.; Kim, Y.-M.; Kim, Y.S.; Choi, E.K.; Kim, D.-Y.; Kim, J.-H.; Nam, J.-H.; Kim, Y.T. Radiation therapy is a treatment to be considered for recurrent epithelial ovarian cancer after chemotherapy. *Tumori J.*, 2011; 97: 590–595. [CrossRef]
 90. Artiga, Á.; Sevilla, I.S.; De Matteis, L.; Mitchell, S.G.; De La Fuente, J.M. Current status and future perspectives of gold nanoparticle vectors for siRNA delivery. *J. Mater. Chem.*, 2019; 7: 876–896. [CrossRef] [PubMed]
 91. Kong, L.; Qiu, J.; Shi, X. Multifunctional PEI-entrapped gold nanoparticles enable efficient delivery of therapeutic siRNA into glioblastoma cells. *J. Control. Release*, 2017; 259: 83–84. [CrossRef]
 92. Fitzgerald, K.A.; Rahme, K.; Guo, J.; Holmes, J.D.; O'Driscoll, C.M. Anisamide-targeted gold nanoparticles for siRNA delivery in prostate cancer - synthesis, physicochemical characterisation and in vitro evaluation. *J. Mater. Chem. B.*, 2016; 4: 2242–2252. [CrossRef]
 93. Labala, S.; Jose, A.; Chawla, S.R.; Khan, M.S.; Bhatnagar, S.; Kulkarni, O.P.; Venuganti, V.V.K. Effective melanoma cancer suppression by iontophoretic co-delivery of STAT3 siRNA and imatinib using gold nanoparticles. *Int. J. Pharm.*, 2017; 52: 407–417. [CrossRef]
 94. Yi, Y.; Kim, H.J.; Zheng, M.; Mi, P.; Naito, M.; Kim, B.S.; Min, H.S.; Hayashi, K.; Perche, F.; Toh, K.; et al. Glucose-linked sub-50-nm unimer polyion complex-assembled gold nanoparticles for targeted siRNA delivery to glucose transporter 1-overexpressing breast cancer stem-like cells. *J. Control. Release*, 2019; 295: 268–277. [CrossRef]
 95. Ahwazi, R.P.; Kiani, M.; Dinarvand, M.; Assali, A.; Tekie, F.S.M.; Dinarvand, R.; Atyabi, F. Immobilization of HIV-1 TAT peptide on gold nanoparticles: A feasible approach for siRNA delivery. *J. Cell. Physiol.*, 2019; 235: 2049–2059. [CrossRef]
 96. Riley, R.S.; Day, E.S. Gold nanoparticle-mediated photothermal therapy: applications and opportunities for multimodal cancer treatment. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.*, 2017; 9: 1449. [CrossRef]
 97. Chen, W.; Zhang, S.; Yu, Y.; Zhang, H.; He, Q. Structural-Engineering Rationales of Gold Nanoparticles for Cancer Theranostics. *Adv. Mater.*, 2016; 28: 8567–8585. [CrossRef] [PubMed]
 98. Wang, J.; Wu, X.; Shen, P.; Wang, J.; Shen, Y.; Shen, Y.; Webster, T.J.; Deng, J. Applications of Inorganic Nanomaterials in Photothermal Therapy Based on Combinational Cancer Treatment. *Int. J. Nanomed.*, 2020; 15: 1903–1914. [CrossRef] [PubMed]

99. Liu, M.; Huang, P.; Wang, W.; Feng, Z.; Zhang, J.; Deng, L.; Dong, A. An injectable nanocomposite hydrogel co-constructed with gold nanorods and paclitaxel-loaded nanoparticles for local chemophotothermal synergetic cancer therapy. *J. Mater. Chem. B*, 2019; 7: 2667–2677. [CrossRef] [PubMed]
100. Song, C.; Li, F.; Guo, X.; Chen, W.; Dong, C.; Zhang, J.; Zhang, J.; Wang, L.; Chen, N. Gold nanostars for cancer cell-targeted SERS-imaging and NIR light-triggered plasmonic photothermal therapy (PPTT) in the first and second biological windows. *J. Mater. Chem. B*, 2019; 7: 2001–2008. [CrossRef]
101. Ortiz de Solorzano, I.; Mendoza, G.; Arruebo, M.; Sebastian, V. Customized hybrid and NIR-light triggered thermoresponsive drug delivery microparticles synthesized by photopolymerization in a one-step flow focusing continuous microreactor. *Colloids Surf. B Biointerfaces*, 2020; 190: 110904. [CrossRef]
102. Gobin, A.M.; Watkins, E.M.; Quevedo, E.; Colvin, V.L.; West, J.L. Near-Infrared-Resonant Gold/Gold Sulfide Nanoparticles as a Photothermal Cancer Therapeutic Agent. *Small*, 2010; 6: 745–752. [CrossRef]
103. Zheng, X.J.; Chow, J.C.L. Radiation dose enhancement in skin therapy with nanoparticle addition: A MonteCarlo study on kilovoltage photon and megavoltage electron beams. *World J. Radiol.*, 2017; 9: 63–71. [CrossRef]
104. Wang, Z. Plasmon—resonant gold nanoparticles for cancer optical imaging. *Sci. China Ser. G Phys. Mech. Astron.*, 2013; 56: 506–513. [CrossRef]
105. Zhao, N.; Pan, Y.; Cheng, Z.; Liu, H. Gold nanoparticles for cancer theranostic - a brief update. *J. Innov. Opt. Health Sci.*, 2016; 9: 1630004. [CrossRef].
106. El-Sayed, I.H.; Huang, X.; El-Sayed, M.A. Surface Plasmon Resonance Scattering and Absorption of anti-EGFR Antibody Conjugated Gold Nanoparticles in Cancer Diagnostics: Applications in Oral Cancer. *Nano Lett.*, 2005; 5: 829–834. [CrossRef]
107. Fan, M.; Han, Y.; Gao, S.; Yan, H.; Cao, L.; Li, Z.; Liang, X.-J.; Zhang, J. Ultrasmall gold nanoparticles in cancer diagnosis and therapy. *Theranostics*, 2020; 10: 4944–4957. [CrossRef] [PubMed]